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Enzymatic Preparation of ^{32}P -Labeled β -L-2', 3'-dd-5' ATP and Its Use as a High-Affinity, Conformation-Specific Ligand for Labeling Adenylyl Cyclases

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**ENZYMATIC PREPARATION OF ^{32}P -LABELED $\beta\text{-L-2',3'-dd-5'ATP}$
AND ITS USE AS A HIGH-AFFINITY, CONFORMATION-SPECIFIC
LIGAND FOR LABELING ADENYLYL CYCLASES**

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Abstract: An enzymatic method was developed for the preparation of unlabeled and [$\beta\text{-}^{32}\text{P}$]-labeled $\beta\text{-L-2',3'-dd-5'ATP}$ from the monophosphate with near quantitative yields. $\beta\text{-L-2',3'-dd-5'ATP}$ was a competitive and potent inhibitor of adenylyl cyclases ($\text{IC}_{50} \sim 30 \text{ nM}$). Upon *uv*-irradiation $\beta\text{-L-2',3'-dd-}[\beta\text{-}^{32}\text{P}]\text{-5'ATP}$ directly crosslinked to a chimeric construct of this enzyme. Data suggest that this is a pre-transition state inhibitor and contrasts with the equipotent $2',5'\text{-dd-3'ATP}$, a post-transition state, noncompetitive inhibitor.

Adenylyl cyclases are a family of membrane-bound enzymes that catalyze the formation of adenosine $3':5'$ -monophosphate (cAMP) from $5'\text{ATP}$. Among their many distinct regulatory mechanisms, it is their inhibition by adenine nucleoside $3'$ -phosphates via a domain referred to as the P-site which we find intriguing. It is a property of all known isoforms of mammalian adenylyl cyclases¹, save possibly the enzyme from sperm, and was originally so designated because of the increased inhibitory potency of ligands containing an intact purine². The most potent inhibitors are adenine nucleoside $3'$ -polyphosphates¹⁻³, which inhibit via a dead-end, noncompetitive mechanism implying that they bind to the enzyme in the configuration for and at the leaving site of cAMP^{4,5}. Whereas inhibition by P-site ligands has been well characterized biochemically and pharmacologically and potent and specific inhibitors of the enzyme have been synthesized, potent agents targeted to the substrate $5'\text{ATP}$ binding conformation have not been identified.

From a family of $\beta\text{-L-Ado-5'-phosphates}$ we developed a facile enzymatic procedure for the preparation of unlabeled- and ^{32}P -labeled analogs of the most potent of these compounds, $\beta\text{-L-}$

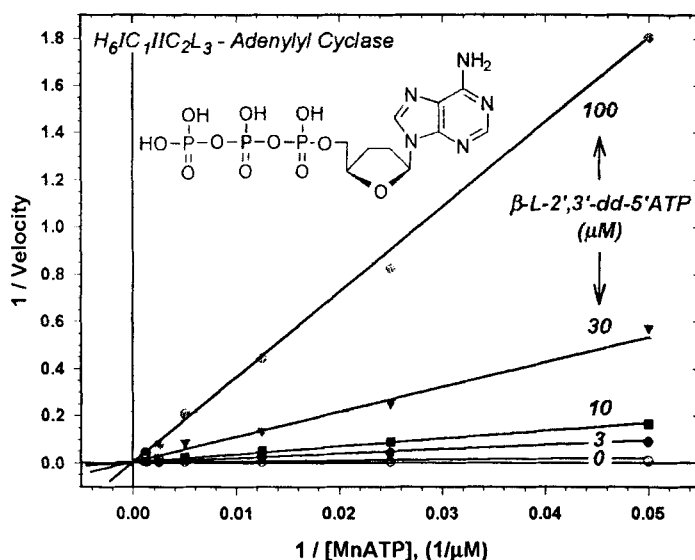


FIGURE 1: Double-reciprocal plot for inhibition of adenylyl cyclase by β -L-2',3'-dd-5'ATP. The chimeric construct $H_6IC_1IIC_2L_3$, comprising the C1 domain of the type I adenylyl cyclase linked with the C2 domain of the type II enzyme⁷, was assayed in the presence of the indicated concentrations of substrate MnATP and β -L-2',3'-dd-5'ATP and 5 mM MnCl_2 fixed in excess of the 5'ATP concentration⁴. Units for velocity are nmols cAMP / (mg protein \cdot min).

2',3'-dd-5'ATP. Unlabeled ligand was prepared from the corresponding 5'-monophosphate upon incubation with myokinase, creatine kinase, and creatine phosphate, with a spiking concentration of 5'ATP. Yields were quantitative. Labeled ligand was similarly prepared, but with a two-step incubation with myokinase and $[\gamma\text{-}^{32}\text{P}]\text{-5'ATP}$ and then a subsequent phosphorylation reaction with creatine phosphate and creatine kinase. Overall yield was ~ 3 mCi of β -L-2',3'-dd- $[\beta\text{-}^{32}\text{P}]\text{-5'ATP}$ from 10 mCi $[\gamma\text{-}^{32}\text{P}]\text{-5'ATP}$. Unlabeled and labeled ligands were purified by sequential ion exchange and ion-pairing reverse phase HPLC.

Inhibition of adenylyl cyclase by β -L-2',3'-dd-5'ATP was competitive with respect to substrate MnATP (Figure 1) and exhibited an $\text{IC}_{50} \sim 30$ nM with native enzyme from rat brain (not shown). This potency compares to that of 2',5'-dd-3'ATP ($\text{IC}_{50} \sim 40$ nM), a non-competitive inhibitor of the enzyme^{1,3,6}. Upon $\alpha\text{-irradiation}$ β -L-2',3'-dd- $[\beta\text{-}^{32}\text{P}]\text{-5'ATP}$ exhibited direct crosslinking to adenylyl cyclase (Figure 2). The best competitive displacement was noted with 2',5'-dd-3'ATP and 2',5'-dd-3'A₄P₆, potent P-site ligands, and 5'AP(CH₂)PP, a substrate analog,

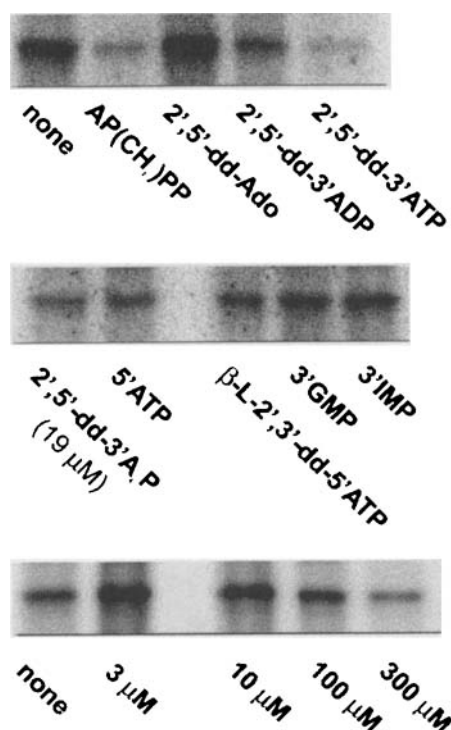


FIGURE 2: Direct photo-crosslinking of β -L-2',3'-dd-[β - 32 P]-5'ATP. Enzyme was irradiated at 300 nm for 20 min in the presence of β -L-2',3'-dd-[β - 32 P]-5'ATP and the indicated other ligands, in a reaction mixture containing 5 mM MnCl_2 , 100 μM forskolin, and 0.5% acetone as sensitizing agent. The top two panels show the effects on labeling of the indicated additional unlabeled ligands, each at 300 μM , except as noted, and the bottom panel shows the effects of unlabeled β -L-2',3'-dd-5'ATP at increasing concentrations. Irradiated enzyme was isolated by denaturing polyacrylamide gel electrophoresis and the extent of crosslinking of ^{32}P -ligand was visualized by PhosphorImager techniques.

but not with 3'GMP, 3'IMP, or 2',5'-dd-Ado. In corroborating experiments β -L-2',3'-dd-[β - ^{32}P]-5'ATP also labeled the enzyme specifically and reversibly in a reversible binding assay, in a manner consistent with an interaction with the catalytically active conformation of the enzyme. The data suggest that β -L-2',3'-dd-5'ATP, and 2',5'-dd-3'ATP interact with adenylyl cyclase at the same site, but with different enzyme conformations. In enzyme structure studies with these nucleotides conformational shifts occurring during catalysis may be determined.

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